Molecular mapping of resistance to Fusarium head blight derived from three *Triticum* species

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INTRODUCTION

Due to yield and quality losses, especially the accumulation of mycotoxins in the grain, *Fusarium* head blight (FHB) remains a serious problem in wheat production. We report here about genetic analysis of FHB resistance derived from three *Triticum* sources: 1) *Triticum macha* (Georgian spelt wheat, 2n=42), 2) *Triticum dicoccum* (cultivated emmer, 2n=28) and 3) *Triticum dicoccoides* (wild emmer, 2n=28). Aim of this work was to unlock novel QTL for diversifying FHB resistance breeding.

MATERIALS AND METHODS

Plant material

Population 1: *T. macha* x Furore, 300 BC₂F_{3:4} lines,

Population 2: *T. dicoccum*-line161 x Helidur, 120 BC₁F₅ lines

Population 3: *T. dicoccum*-line 161 x Floradur, 120 BC₁F₅ lines

Population 4: *T. dicoccoides* – Mt.Gerizim#36 x Helidur, 105 BC_1F_5 lines

Resistance evaluation

Populations 1, 2 and 3 were field evaluated for FHB severity using spray inoculations applying *F. graminearum* or *F. culmorum* conidia suspensions in 7 (or 8) experiments during 4 seasons similarly as described by Buerstmayr et al. (2003). Population 4 was single floret inoculated in one open field and four greenhouse experiments using *F. graminearum* similar to Gladysz et al. (2007).

Genotyping and QTL mapping

The populations were genetically analysed using genome wide SSR and AFLP markers, linkage maps were constructed using *CarthaGène* (de Givry et al. 2004) and QTL analysis was done in *Qgene* (Nelson 1997). with simple and composite interval mapping.

RESULTS AND DISCUSSION

Table 1: QTL estimates for mean FHB severity (means over 7 experiments) in **population 1** (*T. macha* x Furore): chromosomal location, logarithm of odds (LOD) and percent of explained variance (VE) by simple interval mapping (SIM).

Chrom.	Marker interval	Resistance source	LOD>3 no of exp.	SIM	
				LOD	VE
2A	Xgwm296A - Xs12m14_5	T. macha	6	6.2	9
2B	Xs23m13_10 - Xgwm200	T. macha	4	4.3	6
2B	Xwmc317 - Xs24m19_6	T. macha	7	7.9	11
5A	Q-locus (spelt)	T. macha	7	18.7	25
5B	Xs19m14 8 - Xs22m75 1	T. macha	4	6.2	9

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Triticum macha 2n = 42

Triticum dicoccoides Mt.Gerizim#36 2n=28

Triticum dicoccum Line 161 2n=28

 Table 2: QTL estimates for mean FHB severity (means over 8 experiments) in population 2 (*T. dicoccum x* Helidur)

Chrom.	Marker interval	Resistance source	LOD>2.5 no of exp.	SIM	
				LOD	VE
4B	RhtB1	T. dicoccum	6	14	41

 Table 3: QTL estimates for mean FHB severity (means over 7 experiments) in population 3 (*T. dicoccum x* Floradur)

Chrom.	Marker interval	Resistance source	LOD>2.5 no of exp.	SIM	
				LOD	VE
3B	Xgwm493 – Xbarc133	Floradur	4	3.1	12
4B	RhtB1 - Xgwm888	T. dicoccum	3	3.4	13
6B	Xwmc398 - Xwmc397	T. dicoccum	2	2.9	11

Table 4: QTL estimates for mean FHB severity (means over 5 experiments) in **population 4** (*T. dicoccoides x* Helidur)

Chrom.	Marker interval	Resistance source	LOD>2.5 no of exp.	SIM	
				LOD	VE
3A	Xgwm1121 - Xgwm720	T. dicoccoides	4	4.8	20
3A	Xgwm2 - Xgwm779	T. dicoccoides	2	4.5	15
6B	Xs13m24_6 - Xgwm626	T. dicoccoides	4	6.4	26

Summary

Several novel QTL for FHB resistance were discovered, all *T. macha* derived QTL are reported here for the first time. Interestingly, the largest QTL in *T. macha* appears associated with the *Q-locus* (spelt ear type) on 5A. In *T. dicoccum* a strong association between the semi dwarf allele *RhtB1b* and increased FHB susceptibility was evident, especially in the Helidur derived population. Resistance to fungal spread in *T. dicoccoides* was reproducibly associated with QTL on chromosomes 3A and 6B. The 6B QTL form *T. dicoccum* and *T. dicoccum* and *T. dicoccum* as the term of term of the term of ter

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